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Screening of Bioactive Compounds and Potential Drug Targets for Cirrhosis Associated Cardiac Hypertrophy

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ABSTRACT

Background and Objective: Cardiovascular diseases (CVDs) are the leading cause of mortality worldwide, claiming the lives of an estimated 17.9 million people per year. Synthetic medicines are prone to toxicity and long-term side effects. Herbal medicine, on the other hand, is typically directed towards assisting the body's natural healing process and can decrease toxicity and increase therapeutic benefits. The objective of study is to quantitatively analyze the bioactive compounds of Solanum album and Solanum nigrum and in silico analysis to identify the protection pathway of cardiac tissues against cirrhosis-related cardiac hypertrophy using selected herbal compounds. Materials and Methods: Crude extracts were prepared with methanol, ethanol, hexane and chloroform solvents by extraction technique (Maceration method). Phytochemical analysis was carried out with a confirmatory test through TLC for terpenoids, alkaloids, flavonoids, phenols and tannins. Plant extracts of Solanum album and Solanum nigrum are characterized by UV-Vis Spectrophotometer (UV) and Fourier-Transform Infrared Spectroscopy (FTIR). In silico molecular docking with RCSB Protein Data Bank is used to get the three-dimensional structures of AGTR1 (PDB101) and PLC (PDB101). Ligand selection is done with the help of Pub Chem Target and ligand optimization is done by Drug Discovery Studio version 21.1.0 software and PYMOL 4.6.0 used for molecular docking analysis. Results: In vitro studies revealed that methanolic and ethanolic extracts of Solanum nigrum and Solanum album are rich sources of active secondary metabolites. Percentage inhibition against standard (ascorbic acid) showed that herbal extracts have excellent radical scavenging activity. The FTIR spectra of Solanum nigrum and Solanum album extracts showed peaks at 3323 and 3685 cm⁻¹ due to stretching of OH group. The IC₅₀ value of 81.746 µg/mL for Solanum nigrum and 74.048 µg/mL for Solanum album is reported. In silico studies highlights that Quercetin has binding energies of -8.4 kcal/mol compared to Angiotensin II (-7.5 kcal/mol) and Phosphatidylinositol 4,5 bisphosphate (-7.3 kcal/mol) with strong molecular binding action mechanism as compared to the conventional drugs. Conclusion: The plant extracts and their components have shown excellent antioxidant activities, in silico results of the study need in vivo pharmacological experiments for further verification to significantly improve the efficiency of compounds under study.

KEYWORDS

Cardiovascular disease, cirrhotic cardiomyopathy, angiotensin II, natural antioxidants, molecular docking, drug targets

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INTRODUCTION

Despite accounting for a quarter of the world's population, South Asia already bears 60% of the worldwide burden of heart disease¹. Cardiovascular disease (CVD) is a major cause of global death and morbidity. South Asians have a disproportionately high prevalence of coronary artery disease (CAD)². Cirrhotic cardiomyopathy is a kind of cardiac dysfunction that comprises lower cardiac contractility with systolic and diastolic failure, as well as the presence of electrophysiological abnormalities, specifically QT prolongation³. Angiotensin (Ang II) exposure to the heart causes pathological and physiological cardiac hypertrophy, respectively, the signal transduction mechanisms for these effects are complex in nature. It is now clear that the activation of Ang type 1 receptors (AT1R) mediates the hypertrophic response. Reduced AT2R and Mas receptor expression and mitochondrial NADPH oxidase 4 activation may cause cardiac hypertrophy⁴.

A clinical condition known as cirrhotic cardiomyopathy affects people with liver cirrhosis and is characterized by an aberrant and muted response to physiological, pathological or pharmacological stress, but is normal during rest due to increased cardiac output and tactility. Up to 50% of cirrhotic patients receiving liver transplants have symptoms of cardiac dysfunction and overt heart failure accounts for 7 to 21% of mortality following orthotropic liver transplantation⁵. Hypertrophy is brought on by either a protracted increase in the mean contractile "stress" (or tension) or, more likely, a sustained rise in the stroke energy expenditure per unit mass of heart muscle⁶. The development of this condition appears to be influenced by cardiac-adrenergic receptor dysfunction and its signaling role⁷.

Herbal formulas are used as supplemental or alternative treatments since they are inexpensive, safe and have no negative side effects. The two most common conditions of dyslipidemia or increased lipids in the blood, are coronary heart disease (CHD) and cardiovascular disease (CVD), which cause atherosclerosis and cardiac arrest⁸. A well-known *in silico* structure-based method adopted extensively in drug development is molecular docking. Without knowing in advance the chemical makeup of other target modulators, docking enables the discovery of new medicinal compounds, the molecular prediction of ligand-target interactions or the delineation of structure-activity correlations (SAR)⁹. The desire to utilize herbal supplements is fueled by several factors the most important of which are their cost-effective therapeutic potential and safety when compared to mainstream contemporary medicines.

The current study attempted to highlight the potential of herbal extracts as possible treatments for Cirrhosis associated cardiac hypertrophy. Through various analyses and predictions, the indicated plant extracts and their components demonstrated antioxidant and inhibitory activities against standards. Based on the results of the *in silico* analysis, Quercetin improves the molecular mechanism of action against conventional drugs (Angiotensin II). The compound employed in this study may be useful in better-designed research and future therapeutic trials in animal models, obstructing pathogenicity's actions.

MATERIALS AND METHODS

Study area: Medicinal plants named *S. album* and *S. nigrum*, were collected from local pansar. The authentication of plant samples was confirmed by the Taxonomist and Herbarium of the National Agricultural Council, Islamabad Pakistan.

Research protocol: Plants were dried under WHO guidelines. Plants were washed and dried under shade for two weeks. The flowers of plants were separated and pulverized in an electric grinder (NIMA Electric Grinder NM-83000). As 10 g of fine flower powdered will be extracted with 100 mL methanol and ethanol, in a conical flask at 37°C for 72 hrs utilizing electrical orbital shaker (VELP Scientifica Srl via Stazione, 1620865 Usmate (MB)-Italy) then filtered through muslin cloth and. The filtrate is then dried on

Res. J. Med. Plants, 18 (1): 24-31, 2024

a rotary evaporator Rocker Scientific Co. Ltd., 11F., No. 402, Sec. 1, Ren'ai Rd., Linkou Dist., New Taipei City 244014, Taiwan (R.O.C.) until all the solvents get evaporated and the only dry extract is left behind. The percentage yield of all the extracts was calculated by using formula¹⁰:

 $Yield (\%) = \frac{Weight of dry extract}{Weight of dry plant} \times 100$

Plant extracts were screened for phytochemicals; 2 mg/mL of plant extract solution was prepared in respective plant solvent (methanol) and was screened for the following phytochemicals. Fourier-Transform Infrared (FTIR) spectroscopy is an effective tool for the chemical analysis of biological material. The mid-IR spectrum is the most widely used in the sample analysis, but the far- and near-IR spectrum also contributes to providing information about the samples analyzed. This study focused on the analysis of FTIR in the mid-IR spectrum. The ability of the extract to scavenge hydrogen peroxide (H_2O_2) was determined. Aliquot of 0.1 mL of extracts (25-400 µg/mL) was transferred into the Eppendorf tubes and their volume was made up to 0.4 mL with 50 mm phosphate buffer (pH 7.4) followed by the addition of 0.6 mL of H₂O₂ solution (2 mM). The reaction mixture was vortexed and after 10 min of reaction time, its absorbance was measured at 230 nm. Ascorbic acid was used as the positive control. The RCSB (The Research Collaboratory for Structural Bioinformatics Protein Data Bank) is used for accession of a target protein. Quercetin and alpha santalol chemical structures were collected from the PubChem substance database, while Angiotensin II and Phophatidylinositol 4,5 bisphosphate chemical structures were retrieved from the Chem spider database. The PYMOL tool was used to convert the MOL SDF format of these ligands to a PDB file. Drug Discovery Studio version 21.1.0 software is used to optimize the PDB coordinates of the target protein and the Quercetin molecule for docking research. Drug Discovery Studio version 21.1.0 and PYMOL 4.6.0 are used for analysis of target active binding sites. Finally, the Auto Dock Vina was used to dock protein and ligand based on an affinity.

RESULTS

Percentage yield: Methanolic extract of *S. nigrum* gave a yield of 12.4% while ethanolic extract of *S. album* showed more yield of 18.3% as shown in Table 1 and 2.

Phytochemical analysis: Ethanolic plant extracts of *S. album* and methanolic plant extract of *S. nigrum* were tested for a total of five phytochemicals i.e. flavonoids, phenols, tannins and terpenoids were positive. All the results are summarized in Table 3.

Thin layer chromatography: The ethanolic extract of the *S. nigrum* plant successfully showed the presence of all tested secondary metabolites. Samples run in the methanol and acetic acid solvent system clearly showed the presence of phenols. The reddish-grey color appeared after the spray of FeCl₃, confirming the presence of phenols in *S. nigrum* plant extract with Rf values 0.68 and 0.65. Samples run

Table 1 [.] Percentage	vields of methanol	and n-hexane of S	olanum niarum
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Plant solvent	Dry wei	ght (g)	Crude v	veight (g)	(%) yield w/w		
	Methanol	 Methanol n-hexane		n-hexane	Methanol n-hexane		
Solanum nigrum	10	10	1.24	0.34	12.4	3.4	

Among two solvents the methanolic extract of Solanum nigrum showed high percentage yield

Table 2: Percentage yield of ethanol and n-hexane of Solanum album

Plant solvent	Dry we	ight (g)	Crude	weight (g)	(%) yield w/w					
	Ethanol	n-hexane	Ethanol	n-hexane	Ethanol	n-hexane				
Santalum album	<i>n album</i> 10 10		1.83	0.34	18.3	3.4				

Among two solvents the ethanolic extract of *Solanum album* showed a high percentage yield



Fig. 1: FTIR spectrum of *Solanum nigrum* represents different peaks at various wavenumbers There are five peaks in the spectrum region which depicts that plant extract contains complex compounds. Fingerprint region give reading at 1025 cm⁻¹ while other regions represent C-H bending, N-H stretching, C = O stretching and O-H stretching

Table 3: Rf values of various phytochemicals in methanolic berries extract of Solanum nigrum plant

1 2	5						
Phytochemicals	Solvent system	Rf value					
Phenols	Methanol: Acetic acid	A = 0.68					
		B = 0.65					
Alkaloids	Ethyl acetate: Chloroform	A = 0.58					
		B = 0.59					
Flavonoids	Ethyl acetate: N-butanol: Water	A = 0.20					
		B = 0.24					
Tannins	Chloroform: Methanol	A = 0.65					

Among all the phytochemicals, solvent system methanol: Acetic acid showed best Rf value

in ethyl acetate and chloroform solvent system for alkaloids. The appearance of a creamy white color on the plate after Mayer's reagent spray confirmed the presence of alkaloids with the following Rf values 0.58 and 0.59 for the *Solanum nigrum* plant. Samples run in ethyl acetate, n-butanol and water solvent systems were tested for flavonoids. The appearance of a green color after the spray of 3% boric acid and 10% oxalic acid showed the presence of flavonoids with Rf values 0.2 and 0.24 for the *Solanum nigrum* plant. Samples run in chloroform and methanol were tested for tannins. The brownish-grey color appeared after the spray of FeCl₃, confirmed the presence of tannins in the extract with Rf values of 0.65 for *S. nigrum* plant extract. All the reults are shown in Table 3.

FTIR analysis: The FTIR spectra of *S. nigrum* plant extract give peaks at 1025, 1679, 2907, 3323 and 3673 cm⁻¹ summarize in Fig. 1. A peak at 3323 cm⁻¹ is due to the O-H group in the pyranose ring and the N-H group in prime. The FTIR absorption spectra of pure *S. nigrum* are shown in Fig. 1.

The FTIR absorption spectra of pure *S. album* are shown in Fig. 2. The FTIR spectra of *S. album* plant extract give peaks at 1027, 1398, 1611, 2916, 3323 and 3685 cm⁻¹. A peak at 1611 is due to the unsaturated ketone group. A peak at 2916 is due to the alkane group. A peak at 3323 cm⁻¹ is due to the O-H group in the pyranose ring and N-H group in prime and a peak at 3685 cm⁻¹ is due to the 0-H stretching alcohol group. All the results are shown in Fig. 2.

Hydrogen peroxide radical scavenging activity: The IC₅₀ value of methanolic fruit extracts of S. *nigrum* and standard ascorbic acid for scavenging of H_2O_2 was 81.746 and 89.462 µg/mL and the IC₅₀ value of



Fig. 2: FTIR spectrum of *Santalum album* represents different peaks at various wavenumbers There are six peaks in the spectrum region which depicts that plant extract contains more complex compounds than *Solanum nigrum*. Fingerprint region give reading at 1027 and 1398 cm⁻¹ while other regions represent C = O bending, N-H stretching, C = O stretching and O-H stretching



Fig. 3: Extract of *Solanum album* and *Solanum nigrum* at lowest concentration as compared to standard This show maximum percentage inhibition as compared to standard (Ascorbic acid). These results justified the good insights for next animal model studies regarding dose concentration that should be less

ethanolic fruit extracts of *S. album* and standard Ascorbic acid for scavenging of H_2O_2 was 74.048 and 86.559 µg/mL as shown in the Fig. 3.

Ethanolic extract of *S. nigrum* shows IC_{50} value of 81.74 µg/mL±1.8^d while methanolic extract of *S. album* shows IC_{50} value of 74.04 µg/mL±1.7^d which justifies the good insight for antioxidant essays regarding less dose concentration for animal studies compared to ascorbic acid which shows the highest IC_{50} value 86.59 µg/mL±1.9^d and 89.462 µg/mL±1.6^d that is not significant to scavange the free radicals.

In silico analysis: Modes of confirmation from highest to lowest poses are shown in Table 4. By comparing the binding affinities among two ligands Quercetin from present study have the highest binding affinities -8.4 kcal/mol as compared to a natural substrate Angiotensin II having a binding affinity of -7.5 kcal/mol.

By comparing the binding affinities, Quercetin from present study has the highest binding affinities -8.4 kcal/mol and as compared to the natural substrate (Phosphatidylinositol 4,5 bisphosphate) having a binding affinity of -7.3 kcal/moL¹¹ shown in Table 5.

Res. J. Med. Plants, 18 (1): 24-31, 2024

Table 4: Binding affinities of Quercetin and Angiotensin II at the active site of surviving

Ligand	lst	2nd	3rd	4th	5th	6th	7th	8th	9th
Angiotensin II	-7.5	-7.0	-6.8	-6.6	-6.5	-6.4	-6.3	6.3	-5.90
Quercetin	-8.4	-8.3	-7.9	-7.9	-7.7	-7.5	-7.4	-7.4	-7.30

Highest (9th) to lowest (1st) modes of conformation with corresponding RMS (root mean square) binding affinities in G (kcal/mol), ⁴Modes of confirmation from highest to lowest poses

Table 5: Binding affinities of Quercetin and Phosphatidylinositol 4,5 bisphosphate at the active site of surviving

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Ligand	lst	2nd	3rd	4th	5th	6th	7th	8th	9th
Phosphatidylinositol 4,5 bisphosphate	-7.3	-7.2	-6.9	-6.8	-5.9	-5.7	-5.6	-5.5	-5.4
Quercetin	8.4	-8.3	-7.9	-7.9	-7.7	-7.5	-7.4	-7.4	-7.3

Highest (9th) to lowest (1st) modes of conformation with corresponding RMS (root mean square) binding affinities in G (kcal/mol), ⁵Highest to lowest modes of confirmation of Phosphatidylinositol 4,5 bisphosphate and Quercetin

DISCUSSION

Ethanolic extract of *S. album* showed best yield of 18.3% as compared to methanolic extract of *S. nigrum* which is 12.4%. Similar results also supported the data that β -santalol and α - santalol were included in the ethanolic extract at the highest level with 19.6 and $16\%^{12}$. The fractions that are obtained from phytochemical analysis revealed the best Rf value from methanol: Acetic acid solvent system in *S. nigrum* for screening of phenol while the Rf value of *S. album* is 0.2 in methanolic: Water solvent system. Data from literature greatly support the same Rf values for *S. nigrum* which showed two spots having Rf value of 0.65 and 0.68¹³.

Phenols are screened as secondary metabolites for methanolic berries extract of *S. nigrum* plant. The amount of phenols that can be extracted depends on the solvent's polarity, indicating that high polarity solvents are ideal for extraction as reported in previous studies¹⁴. *Solanum nigrum* samples were screened with an FTIR spectrophotometer in the range of 500-4000 cm⁻¹ by solution casting method to determine the chemical interaction of each sample¹⁵.

A functional group was found in similar tests, with absorbance peaks at 3,436, 2,919, 1,606, 1,386, 1,041 and 545 cm⁻¹ corresponding to -OH, -CH2, -COOH, C-C, C-C and C-H¹⁶. The FTIR absorption spectra of pure *S. album* was in the range of 500-4000 cm⁻¹ by solution casting method to determine the chemical interaction of each sample. Similar studies were reported regarding C = O stretching ,C-H stretching at 2916¹⁷. The C-H methylation, also known as the "Magic Methyl Effect", has attracted a lot of attention recently due to its biological significance in medicinal chemistry¹⁸. Ethanolic fruit extracts of *S. nigrum* and methanolic extract of *S. album* showed the best IC₅₀ value of 81.74 and 74.04 µg/mL as compared to standard Ascorbic acid (86.59 and 89.462 µg/mL) for scavenging free radicals. Extracts were capable of scavenging hydrogen peroxide in a concentration-dependent manner¹⁹. Similar studies also supported the *S. nigrum* leaf extracts to scavenge free redicals²⁰. *Solanum nigrum* at its lowest concentration shows maximum percentage inhibition as compared to standard Ascorbic acid²¹.

By comparing the results it is assumed that Quercetin from present study have the highest binding affinity of -8.4 kcal/mol as compared to a natural substrate Phosphatidylinositol 4,5 bisphosphate and Angiotensin II having a binding affinity of -7.3 and -7.5 kcal/mol. According to *in silico* research, AT1R blockade has been demonstrated to inhibit the regression of Ang II-induced cardiac hypertrophy²². Kang *et al.*²³ reported that Quercetin binds with AT1R blockers, like losartan, to effectively reduce cardiac hypertrophy²³. Though the precise causes of the shift from Ang II-induced physiological to pathological cardiac hypertrophy remain unclear, one of the most significant pathogenic elements thought to be involved in this process could be the establishment of a critical level of oxidative stress. To verify quercetin's effectiveness with ATIR blockers, more *in vivo* research could be conducted.

CONCLUSION

The current study attempted to highlight the potential of herbal extracts as possible treatments for Cirrhosis associated cardiac hypertrophy. Through various analysis and *in silico* predictions, the indicated plant extracts and their components demonstrated antioxidant activities. Based on the results of the Insilico analysis, Quercetin may improve the molecular mechanism of action against conventional drugs targeting Angiotensin II which needs to be verified by *in vivo* study.

SIGNIFICANCE STATEMENT

The quantitative and qualitative analysis of bioactive compounds revealed the therapeutic properties and presence of different types of important functional groups in *S. album* and *S. nigrum*. *In vitro* studies revealed that methanolic and ethanolic extracts of both plants yield active secondary metabolites with excellent radical scavenging activity. With the help of *in silico* analysis, unique protection pathways have been identified against cirrhosis-related cardiac hypertrophy using a selected herbal compound, quercetin which may improve the molecular mechanism of action against conventional drug target Angiotensin II. The compound employed in this study may be useful in better understanding of designing targeted drugs for future therapeutic trials in animal models.

REFERENCES

- 1. Martinez-Amezcua, P., W. Haque, R. Khera, A.M. Kanaya and N. Sattar *et al.*, 2020. The upcoming epidemic of heart failure in South Asia. Circ.: Heart Fail., Vol. 13. 10.1161/CIRCHEARTFAILURE.120.007218.
- 2. Fernando, E., F. Razak, S.A. Lear and S.S. Anand, 2015. Cardiovascular disease in South Asian migrants. Can. J. Cardiol., 31: 1139-1150.
- 3. Møller, S., J.D. Hove, U. Dixen and F. Bendtsen, 2012. New insights into cirrhotic cardiomyopathy. Int. J. Cardiol., 167: 1101-1108.
- 4. Bhullar, S.K. and N.S. Dhalla, 2022. Angiotensin II-induced signal transduction mechanisms for cardiac hypertrophy. Cells, Vol. 11. 10.3390/cells11213336.
- 5. Zardi, E.M., A. Abbate, D.M. Zardi, A. Dobrina and D. Margiotta *et al.*, 2010. Cirrhotic cardiomyopathy. J. Am. Coll. Cardiol., 56: 539-549.
- 6. Badeer, H.S., 1972. Pathogenesis of cardiac hypertrophy in coronary atherosclerosis and myocardial infarction. Am. Heart J., 84: 256-264.
- 7. Myers, R.P. and S.S. Lee, 2000. Cirrhotic cardiomyopathy and liver transplantation. Liver Transpl., 6: S44-S52.
- 8. Sharma, R. and A. Itharat, 2017. Herbal supplements or herbs in heart disease: Herbiceutical formulation, clinical trials, futuristic developments. J. Cardiol. Cardiovasc. Ther., Vol. 3. 10.19080/JOCCT.2017.03.555603.
- 9. Pinzi, L. and G. Rastelli, 2019. Molecular docking: Shifting paradigms in drug discovery. Int. J. Mol. Sci., Vol. 20. 10.3390/ijms20184331.
- 10. Liu, X., Q. Zhang, Z. Hong and D. Xu, 2022. Induction of heartwood formation in young Indian sandalwood (*Santalum album* L.) by gas elicitors. Front. Plant Sci., Vol. 13. 10.3389/fpls.2022.961391.
- Adeoye, A.O., J.O. Olanlokun, H. Tijani, S.O. Lawal, C.O. Babarinde, M.T. Akinwole and C.O. Bewaji, 2019. Molecular docking analysis of apigenin and quercetin from ethylacetate fraction of *Adansonia digitata* with malaria-associated calcium transport protein: An *in silico* approach. Heliyon, Vol. 5. 10.1016/j.heliyon.2019.e02248.
- 12. Sharifi-Rad, J., C. Quispe, A. Turgumbayeva, Z. Mertdinç and S. Tütüncü *et al.*, 2023. *Santalum* Genus: Phytochemical constituents, biological activities and health promoting-effects. Zeitschrift Naturforsch. C, 78: 9-25.
- 13. Goel, K., R. Singh, V. Saini and M. Sharma, 2022. Physicochemical evaluation of *Solanum nigrum* Linn. and *Tribulus terrestris* Linn. Int. J. Health Sci., 6: 10786-10795.

- 14. Staveckienė, J., J. Kulaitienė, D. Levickienė, N. Vaitkevičienė and V. Vaštakaitė-Kairienė, 2023. The effect of ripening stages on the accumulation of polyphenols and antioxidant activity of the fruit extracts of *Solanum* species. Plants, Vol. 12. 10.3390/plants12142672.
- 15. Cao, J., C. Wang, Y. Zou, Y. Xu and S. Wang *et al.*, 2023. Colorimetric and antioxidant films based on biodegradable polymers and black nightshade (*Solanum nigrum* L.) extract for visually monitoring *Cyclina sinensis* freshness. Food Chem.: X, Vol. 18. 10.1016/j.fochx.2023.100661.
- Mohanaparameswari, S., M. Balachandramohan, P. Sasikumar, C. Rajeevgandhi and M. Vimalan *et al.*, 2023. Investigation of structural properties and antibacterial activity of ago nanoparticle extract from *Solanum nigrum/Mentha* leaf extracts by green synthesis method. Green Process. Synth., Vol. 12. 10.1515/gps-2023-0080.
- Umdale, S., M. Ahire, V. Aiwale, A. Jadhav and P. Mundada, 2020. Phytochemical investigation and antioxidant efficacy of wild, underutilized berries of economically important Indian Sandalwood (*Santalum album* L.). Biocatal. Agric. Biotechnol., Vol. 27. 10.1016/j.bcab.2020.101705.
- 18. Mostafa, M.M.M., T.S.S. Saleh and N.S.I. Ahmed, 2023. The role of catalysts in functionalization of C-H and C-C bonds. Catalysts, Vol. 13. 10.3390/catal13020377.
- 19. Chen, J.F., S.W. Wu, Z.M. Shi, Y.J. Qu, M.R. Ding and B. Hu, 2023. Exploring the components and mechanism of *Solanum nigrum* L. for colon cancer treatment based on network pharmacology and molecular docking. Front. Oncol., Vol. 13. 10.3389/fonc.2023.1111799.
- 20. D.A. Martin G., J.C.P. Maldonado and O.E.C. González, 2022. HPLC-DAD analysis, antifungal and antioxidant activity of *Solanum dolichosepalum* bitter extracts and fractions. Braz. J. Pharm. Sci., Vol. 58. 10.1590/s2175-97902022e20350.
- 21. Yimer, A., S.F. Forsido, G. Addis and A. Ayelign, 2023. Phytochemical profile and antioxidant capacity of some wild edible plants consumed in Southwest Ethiopia. Heliyon, Vol. 9. 10.1016/j.heliyon.2023.e15331.
- 22. Lee, C.Y., H.K. Park, B.S. Lee, S. Jeong and S.A. Hyun *et al.*, 2020. Novel therapeutic effects of pterosin B on ang II-induced cardiomyocyte hypertrophy. Molecules, Vol. 25. 10.3390/molecules25225279.
- 23. Kang, B.Y., J.A. Khan, S. Ryu, R. Shekhar, K.B. Seung and J.L. Mehta, 2010. Curcumin reduces angiotensin II-mediated cardiomyocyte growth via LOX-1 inhibition. J. Cardiovasc. Pharmacol., 55: 176-183.